

ORIGINAL ARTICLE

Evaluation of a polyherbal formulation for the management of wet litter in broiler chickens: Implications on performance parameters, cecal moisture level, and footpad lesions

Saravanakumar Marimuthu¹, Brindhalakshmi Balasubramanian¹, Ramasamy Selvam², Prashanth D'Souza¹

¹Animal Health Science, R&D Centre, Natural Remedies Private Limited, Veerasandra Industrial Area, Bengaluru 560 100, India

²Technocommercial, Marketing, Natural Remedies Private Limited, Veerasandra Industrial Area, Bengaluru 560 100, India

ABSTRACT

Objective: The study was carried out to develop a wet litter model with magnesium chloride to assess the effectiveness of a polyherbal formulation (PHF) on growth performance, litter and cecal moisture (LCM) level, cecal consistency (CC) score, and footpad lesions (FPLs) score in Ross 308 broiler chickens.

Materials and Methods: 1,200 one-day-old chicks were assigned into five groups: normal control, negative control [NTC; treated with 1.7% magnesium chloride hexahydrate ($MgCl_2 \cdot 6H_2O$)], and three treatment groups, T1, T2, and T3, where 750, 1,000, and 2,000 gm/ton of PHF, respectively, were supplemented. All the groups were fed a basal diet until day 7. However, the NTC and treatment groups were fed a diet with $MgCl_2$ from days 8 to 42.

Results: The addition of $MgCl_2$ for 35 days worsened the growth performance traits in broilers and induced wet litter problems (FPL, high LCM, and poor CC) in the NTC group. However, PHF (750, 1,000, and 2,000 gm/ton) ameliorated the negative effect of a diet with $MgCl_2$ on growth performance and wet litter problems, but a better response with respect to LCM and CC was observed in 2,000 gm/ton of PHF group, followed by that in 1,000 gm/ton of PHF group and 750 gm/ton of PHF group on day 42.

Conclusion: The wet litter broiler model was developed through excessive feeding of $MgCl_2$, which caused the performance parameters to worsen and the emergence of problems associated with the wet litter. Supplementation with PHF ameliorated these problems and, therefore, it can be used for the management of wet litter in poultry.

ARTICLE HISTORY

Received July 11, 2019

Revised September 30, 2019

Accepted October 01, 2019

Published October 30, 2019

KEYWORDS

Chickens; magnesium chloride; polyherbal formulation; wet litter



This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 Licence (<http://creativecommons.org/licenses/by/4.0>)

Introduction

Deep litter system plays a key role in maintaining the bird's health, comfort, welfare, and production efficiency. Typically, litter materials (wood shavings, sawdust, peanut hulls, and rice husk) are continuously blended with spilled food, feathers, and poultry fecal waste, preventing the production of harmful pathogens in the poultry house [1,2]. Furthermore, it acts as a moisture absorber, but the capacity varies from 15% to 45% based on the type of litter material used in the shed [3,4]. However, the moisture-absorbing capacity of litter is completely saturated when the

moisture content of chicken droppings (watery/loose) increases owing to the high water intake. As a result, the addition of water surpasses the removal of water (evaporation), leading litter moisture problems in the poultry sheds. Wet litter is the complex issue caused by multiple interrelated factors, such as management and housing of birds [5,6], disease and diet control [7], and gut health factors [8]. Additionally, it is one of the most significant factors that contribute to the induction of footpad lesions (FPLs), also called footpad dermatitis (FPD), which has become a critical concern in contemporary chicken meat production.

Correspondence Saravanakumar Marimuthu ✉ saravana.k@naturalremedy.com 📧 Animal Health Science, Research and Development Centre, Natural Remedies Private Limited, Veerasandra Industrial Area, Bengaluru 560 100, India.

How to cite: Marimuthu S, Balasubramanian B, Selvam R, D'Souza P. Evaluation of a polyherbal formulation for the management of wet litter in broiler chickens: Implications on performance parameters, cecal moisture level, and footpad lesions. *J Adv Vet Anim Res* 2019; 6(4):536–543.

This is supported by several authors [3,9,10] who observed a high incidence of FPD associated with high litter moisture concentrations in broiler sheds [9,11]. Furthermore, it was demonstrated that the high prevalence of FPD reduced the growth performance (weight gains by 7.75% and feed conversion efficiency by 4.16%) of 37-day-old meat-type chickens [12]. Therefore, attention must be given to the wet litter problem and it should be addressed to prevent production losses and uphold animal health and welfare.

Wet droppings can be effectively rectified by identifying the underlying causal factors, for instance, the management of drinkers and proper shed ventilation would prevent problems due to environmental or housing factors. Similarly, any nutritional disturbance alters the excreta quality of meat chickens, which can be improved by influencing the microbial phytase and dietary electrolyte balance in the feed [13,14]. Similarly, the addition of antibiotics at subtherapeutic concentrations reduces the prevalence of coccidiosis and necrotic enteritis infection, which is the most frequently cited condition among the relevant diseases. However, with the ban of in-feed antibiotics and zinc oxide, well-known anti-diarrheal agents in piglets [15], in European countries animal health deteriorated, which include increased loose droppings/diarrhea due to gut disturbances, weight loss, and high mortality of early post-weaning pigs and broilers [16,17]. Hence, there is an unmet need for natural alternatives to prevent the wet droppings, and as far as we know, no literature related to the effect of natural feed additives on the moisture of broilers excreta is published. The polyherbal formulation (PHF) under investigation, Stodi[®], is a blend of several Indian medicinal plants (*Punica granatum*, *Andrographis paniculata*, *Acacia nilotica*, *Terminalia bellirica*, and *Holarrhena antidysenterica*) traditionally known and used individually to manage diarrheal disorders. However, there are no scientific reports that demonstrate their anti-diarrheal activity in a combined form in a wet litter model in broilers.

Numerous models are available to evaluate wet litter problems in broiler chickens. One of them is that the addition of minerals in the diet at high doses can induce loose droppings/diarrhea in broilers by altering osmolarity and water reabsorption in the gastrointestinal tract (GIT), but the effect varies depending on individual minerals [18]. Magnesium, an indispensable positively charged ion in the diet of most animals [19], is commonly used as a laxative in humans [20,21] and as an osmoregulators in animal diarrhea models [22–25]. It has been reported that magnesium sulfate alters the luminal osmotic pressure, preventing the reabsorption of water from the digesta, and increasing the excreta moisture [25]. Similarly, higher levels of dietary magnesium oxide reduced the transit time of intestinal digestion in broilers [26].

Magnesium chloride ($MgCl_2$) is less toxic [27] and has the capacity to increase the excreta moisture content in comparison with other magnesium sources [28]. Therefore, it was used in the present study to increase the digesta moisture and reduce the water reabsorption instead of increasing water consumption. The objective of the study was to induce wet litter problems using a diet with $MgCl_2$ and evaluate the effect of PHF on growth performance, litter moisture, cecal moisture and consistency, and footpad lesion in Ross 308 broilers.

Materials and Methods

Polyherbal formulation

Stodi[®], a registered PHF formulated by Natural Remedies Pvt. Ltd., Bengaluru, India. It contains rinds of *P. granatum*, aerial parts of *A. paniculata*, bark of *A. nilotica*, fruits of *T. bellirica*, and bark of *H. antidysenterica*.

Experimental setup and ethical approval

The experimental setup and the management of birds (Table 1) were followed as per Selvam et al. [29]. The present study was performed in compliances with guidelines laid down for the care and the use of animals, and protocol approval was provided by the Institutional Ethics Committee [No. AHS/PR/01/2018].

Experimental plan and feeding level

Twelve hundred Ross 308 chicks (1-day-old) were supplied by Kavi Protein and Feed Pvt. Ltd. (Bengaluru) and were allocated equally into five groups (six replicates/group; 40 birds/replicate), consisting of normal control (NC), negative control [NTC; treated with 1.7% magnesium chloride hexahydrate ($MgCl_2 \cdot 6H_2O$)], and three treatment groups, T1, T2, and T3, where 750, 1,000, and 2,000 gm/ton of PHF, respectively, were supplemented. All groups were fed with a basal diet (Table 2) until day 7. However, the NTC and treatment groups were fed with a customized diet (1.7% $MgCl_2$) from day 8 to day 42.

Assessment of zootechnical parameters

The mortality of chicks was observed in each pen, once daily for 42 days. The chicks were weighed individually on days 1, 7, 21, 28, 35 and, finally, on 42nd day. The quantity of feed consumed by birds was determined on 7th, 21st, 28th, 35th, and 42nd days of the experiment by deducting the quantity of remaining feed over the total quantity of feed offered per pen, and feed efficiency (feed conversion ratio, FCR) was calculated by the ratio between the total quantity of feed consumed and the total body weight gain.

Footpad lesion

Footpad lesion was scored using a four-point grading scale [30] on days 35 and 42. It was allocated to one of four classes: score 0, normal footpad; score 1, <25% area covered by lesions; score 2, 25%–50% area covered by lesions; and score 3, >50% area covered by lesions. The ratio of cumulative lesion scores to that of 48 arbitrarily picked birds was considered as a mean footpad lesion score.

Cecal consistency

Cecal consistency (CC) was scored for six birds in each group on day 42. Briefly, the consistency was assigned to one of four classes: score 1, normal (semi-solid and well-formed); score 2, sticky droppings (gelatinous); score 3, mild diarrhea (fluid feces and not well formed); and score 4, severe diarrhea (watery and projectile).

Litter and cecal moisture content

Litter samples were taken from five different locations per pen (replicate) and pooled before measurement on days 21, 28, 35, and 42. The moisture content (%) was evaluated by weight loss on drying using a Hot Air Oven (FS-405 Model) purchased from Advantec Co., Ltd., (Saijyo, Japan). In summary, the pooled litter sample was weighed and allowed to dry at 105°C for 24 h. The dried sample was reweighed, and the weight difference (before and after drying) constitutes the moisture content. A similar procedure was used to evaluate the moisture content (%) in the cecal samples ($n = 6/\text{group}$) on day 42.

Statistical analysis

The statistical analysis was performed as per the procedures described by Selvam et al. [29], and $p < 0.05$ was considered as statistically significant.

Results

Zootechnical parameters of chickens

The initial live body weight in all the groups was not significantly different, indicating the uniform distribution of the chicks for each pen on day 1. The MgCl_2 diet group displayed a reduction in body weight gain on days 35 and 42. However, all the treatment groups (T1, T2, and T3) had significantly increased the body weight gain on days 21, 28, 35, and 42 (Table 3). Similarly, FCR was significantly worsened in the NTC compared to that in the NC group on days 21, 28, 35, and 42. Nevertheless, the groups treated with PHF showed a numerical improvement in the FCR on days 28, 35, and 42 (Table 4).

Footpad lesion score

Footpad lesion score was found to be high in the NTC group as compared to the NC group on days 35 and 42. However, the PHF-treated groups (750, 1,000, and 2,000 gm/ton) showed a numerical improvement in the footpad lesion score compared to that in the NTC group on days 35 and 42 (Table 5).

Litter moisture content

Litter moisture (gm/100 gm) content was significantly increased in the NTC group when compared to the NC group throughout the experimental period. However, litter moisture content (gm/100 gm) was numerically improved in the PHF-treated groups (750, 1,000, and 2,000 gm/ton) compared to that in the NTC group on days 35 and 42 (Table 6).

Cecal consistency score and cecal moisture content

CC score and moisture content (gm/100 gm) of NTC group were worsened significantly as compared to the NC group, whereas these parameters improved after PHF supplementation with the best response being observed in the 2,000 gm/ton of PHF group compared to that observed in the NTC group on day 42 (Table 7).

Discussion

In general, the functions of litter materials are to provide insulation to the bird by creating cushions on the hard floor and facilitate the evaporation of broiler excreta and spilled water by absorbing the moisture in the poultry house. However, any physiological disturbances due to the electrolyte imbalance, non-starch polysaccharides diets, and environmental change upsurge the water intake by stimulating the hypothalamus, which further dilutes the blood and suppresses the secretion of anti-diuretic hormone. This, in turn, urges the high urine output, and to compensate for

Table 1. Environmental conditions.

| Day | Temperature at chick level (°C) | Relative Humidity (%) | Photoperiod |
|-------|---------------------------------|-----------------------|-----------------------------|
| 1–3 | 29–30 | 60–70 | |
| 4–7 | 28–30 | 50–60 | 23 h light and 1 h darkness |
| 8–9 | 27–28 | 50–60 | 22 h light and 2 h darkness |
| 10–11 | 27–28 | 50–60 | 21 h light and 3 h darkness |
| 12–26 | 25–27 | 50–60 | |
| > 27 | 24–26 | 50–60 | 20 h light and 4 h darkness |

Table 2. Feed Composition (kg/ton).

| Type of Feed | Pre-starter | Starter | Finisher |
|--|-------------|-------------|-------------|
| | (Day 1–10) | (Day 11–24) | (Day 25–42) |
| Corn local | 550.00 | 595.40 | 580.00 |
| Rice bran | 24.50 | - | 45.24 |
| Soya bean meal (46.5%) | 340.00 | 290.00 | 274.00 |
| Corn gluten meal (60%) | - | 19.60 | - |
| ^a Deoiled rice bran/MgCl ₂ | 17.00 | 17.00 | 17.00 |
| Limestone fine | 11.00 | 9.00 | 8.00 |
| Dicalcium phosphate | 14.50 | 14.50 | 12.00 |
| Salt common | 2.30 | 2.50 | 2.80 |
| Sodium bicarbonate | 3.00 | 3.20 | 2.60 |
| Blended oil (Veg) | 21.00 | 33.00 | 44.00 |
| L -Threonine 98.5% | 1.10 | 1.00 | 0.80 |
| L-Lysine | 3.00 | 2.80 | 1.65 |
| DL-Methionine 99% | 3.40 | 2.90 | 2.60 |
| Choline 60% | 0.65 | 0.60 | 0.50 |
| ^b Trace mineral mixture broiler | 1.20 | 1.20 | 1.50 |
| Vitamin C | - | 0.25 | 0.25 |
| Betaine | 0.75 | 0.75 | 0.75 |
| Toxin binders | 1.00 | 1.00 | 1.00 |
| Organic selenium | 0.09 | 0.06 | 0.06 |
| Sodium butyrate | 0.50 | 0.25 | 0.25 |
| ^c Premix broiler | 5.00 | 5.00 | 5.00 |
| Metabolizable Energy Kcal | 3,000 | 3,125 | 3,150 |
| Crude Protein % | 21 | 20 | 18.5 |

^aDeoiled rice bran (DORB) was replaced by Magnesium chloride hexahydrate (MgCl₂·6H₂O)

^bMineral premix supplied the following per kilogram: Fe, 40 gm; Cu, 10 gm; Mn, 100 gm; Zn, 100 gm; Se, 0.25 gm; and I, 1.5 gm.

^cPremix broiler composition (5 kg) supplied the following per kilogram: antioxidants, 0.125 kg; emulsifier, 0.500 kg; Phytase, 0.100 kg; acidifier, 1.000 kg; liver tonic, 0.500 kg; ^dvitamin premix, 0.500 kg; DORB, 2.275 kg.

^dVitamin premix supplied the following per kilogram of vitamin premix: vitamin A, 25 MIU; vitamin D₃, 5 MIU; vitamin E, 24 IU; vitamin K, 3 gm; vitamin B₁, 3 gm; vitamin B₂, 10 gm; vitamin B₆, 4 gm; vitamin B₁₂, 0.015 gm; niacin, 30 gm; pantothenic acid, 20 gm; folic acid, 1 gm.

this loss, birds consume more water and excrete the watery droppings. As a result, the litter becomes damp, causing the development of FPD as well as worsening the production performance of chickens. Considering the wet droppings/wet litter as a useful early warning sign, the underlying cause must be identified and addressed as a matter of urgency to maintain the optimum health of birds.

Since the inclusion of excessive Mg leads to skeletal abnormalities in young birds [19], broiler chickens were raised on a diet with MgCl₂ (1.7%) from day 8 to day 42 in the current study. Birds raised on a diet with MgCl₂ for 35 days gained lesser body weight gain (1.7%) with higher feed intake (90 gm/unit of body weight) than birds in the NC group. These results are supported by several authors

[28,31,32] who have observed the limited toxic effect of Mg on broiler performance. Similarly, CC and cecal and litter moisture content were worsened in chickens fed a diet with MgCl₂. This is consistent with the observations of [28], who reported that birds excreted the droppings with high moisture content when they were treated with three different Mg sources. Magnesium, a divalent cation, alters the osmolarity of the digesta [18] due to its poor absorbable character [21] and produces a laxative effect [21,33]. However, other authors did not observe any difference in the intestine even after increasing Mg from 0.15% to 0.80%, suggesting that the endocrine hormonal and neuronal effects were responsible for Mg-mediated diarrhea in broiler chickens. Nevertheless, the high concentration of

Table 3. Effect of PHF on body weight (gm) in Ross 308 broiler chickens (MgCl₂ induced wet litter).

| Group | Day 1 | Day 7 | Day 21 | Day 28 | Day 35 | Day 42 |
|--------------------------|--------------|---------------|----------------------------|-------------------------------|-------------------------------|--------------------------------|
| NC | 46.87 ± 0.22 | 129.20 ± 1.19 | ^a 726.62 ± 7.28 | ^a 1,185.95 ± 12.73 | ^a 1,829.75 ± 20.44 | ^{ab} 2,494.75 ± 28.89 |
| NTC | 47.15 ± 0.24 | 135.08 ± 0.95 | ^a 736.86 ± 6.56 | ^a 1,188.28 ± 12.68 | ^a 1,783.08 ± 19.73 | ^a 2,452.91 ± 29.13 |
| NTC + PHF (750 gm/ton) | 46.72 ± 0.24 | 137.88 ± 1.15 | ^b 771.53 ± 9.00 | ^b 1,261.65 ± 14.30 | ^b 1,886.60 ± 22.79 | ^b 2,536.90 ± 31.51 |
| NTC + PHF (1,000 gm/ton) | 47.44 ± 0.22 | 139.40 ± 1.30 | ^b 789.13 ± 7.77 | ^b 1,282.73 ± 13.88 | ^b 1,898.48 ± 22.37 | ^b 2,534.35 ± 32.43 |
| NTC + PHF (2,000 gm/ton) | 46.86 ± 0.24 | 138.44 ± 1.14 | ^b 773.58 ± 9.06 | ^b 1,264.57 ± 13.48 | ^b 1,888.78 ± 21.54 | ^{ab} 2,525.14 ± 31.61 |

Values are expressed as mean ± standard error mean (SEM); *n* = 226–238; ^{a–b}Means bearing different superscripts were significantly different (*p* < 0.05) by one-way analysis of variance (ANOVA) with location as a blocking factor followed by least significance difference (LSD) test using Statistical Package for the Social Sciences (SPSS).

Table 4. Effect of PHF on FCR in Ross 308 broiler chickens (MgCl₂ induced wet litter).

| Group | Day 7 | Day 21 | Day 28 | Day 35 | Day 42 |
|--------------------------|--------------|---------------------------|----------------------------|---------------------------|---------------------------|
| NC | 0.859 ± 0.02 | ^a 1.526 ± 0.01 | ^a 1.682 ± 0.02 | ^a 1.752 ± 0.01 | ^a 1.812 ± 0.01 |
| NTC | 0.856 ± 0.01 | ^b 1.579 ± 0.02 | ^b 1.765 ± 0.02 | ^b 1.848 ± 0.02 | ^b 1.902 ± 0.02 |
| NTC + PHF (750 gm/ton) | 0.856 ± 0.02 | ^b 1.577 ± 0.02 | ^c 1.728 ± 0.01 | ^b 1.819 ± 0.01 | ^b 1.883 ± 0.01 |
| NTC + PHF (1,000 gm/ton) | 0.858 ± 0.01 | ^b 1.586 ± 0.01 | ^{bc} 1.744 ± 0.01 | ^b 1.833 ± 0.01 | ^b 1.891 ± 0.01 |
| NTC + PHF (2,000 gm/ton) | 0.857 ± 0.01 | ^b 1.584 ± 0.02 | ^{bc} 1.744 ± 0.01 | ^b 1.834 ± 0.02 | ^b 1.893 ± 0.02 |

Values are expressed as mean ± SEM; *n* = 6; ^{a–c}Means bearing different superscripts were significantly different (*p* < 0.05) by One-way ANOVA with location as a blocking factor followed by LSD using SPSS.

Table 5. Effect of PHF on footpad lesion.

| Group | Day 35 | Day 42 |
|--------------------------|--------|--------|
| NC | 0.0 | 0.0 |
| NTC | 1.0 | 1.0 |
| NTC + PHF (750 gm/ton) | 0.9 | 0.6 |
| NTC + PHF (1,000 gm/ton) | 0.8 | 0.5 |
| NTC + PHF (2,000 gm/ton) | 0.8 | 0.8 |

Values are expressed as mean.

both Mg and Cl ions due to MgCl₂ inclusion, was excreted from the kidneys and disturbs the osmotic value after reaching the distal GIT [34]. This, in turn, prevents the water reabsorption and increases the digesta moisture, which is confirmed by high cecal moisture contents in the current study. It has also been reported that the high moisture content in the bedding material could significantly raise the incidence of FPD in broilers [35,36], and it was confirmed in the present study as well. The above findings revalidate the hypothesis that excess MgCl₂ can be added in the diet to develop the wet litter model, which is used to evaluate the effect of anti-diarrheal products or agents for the management of wet litter problems in broilers.

Birds supplemented independently with 750, 1,000, and 2,000 gm/ton of PHF showed an improvement in the body weight gain by about 3.4%, 3.3%, and 2.9%, respectively, with better FCR, namely, 19, 11, and 10 gm, respectively, lesser feed per unit body weight gain. In addition,

the weight gain response to PHF (750, 1,000, and 2,000 gm/ton) was greater than that of the NC response. Our results are consistent with some previous studies, which showed that supplementing *A. paniculata* and *A. nilotica* in the diets of broilers increased the growth performance [29,37]. Moreover, PHF improve the cecal consistency and the cecal and litter moisture, which could be due to the presence of *P. granatum*, *A. paniculata*, *A. nilotica*, and *T. bellirica*. Moreover, *P. granatum* is used to treat stomach disorders and diarrheal cases [38] as its different concentrations of the extract ameliorated the diarrhea induced by castor oil by exerting the beneficial effects on GIT in rats [39]. *A. paniculata* has shown a highly significant anti-secretory activity [40] and an anti-diarrheal property against *E. coli* enterotoxins in guinea pigs and rabbit [41]. *A. nilotica* and *T. bellirica* extract reduced the diarrhea induced by castor oil in rats [42,43]. In addition, tannin is one of the chemical constituents present in each herb [44–48], which is used to treat acute diarrhea [49,50]. Recent studies also indicated that tannic acid improved the mucosal resistance [51] and can protect cells from oxidative damage [52,53]. Similarly, the inclusion of PHF was effective in normalizing the increased footpad lesion score caused by a diet with MgCl₂, which could be due to amoebicidal, anti-inflammatory, and analgesic properties of *H. antidysenterica* [54–56]. The findings of the previous literature on individual herbs related to diarrhea confirm that PHF under investigation has the potential to prevent/manage the gut health/wet litter problems.

Table 6. Effect of PHF on litter moisture content (gm/100 gm).

| Group | Day 21 | Day 28 | Day 35 | Day 42 |
|--------------------------|---------------------------|---------------------------|---------------------------|---------------------------|
| NC | ^a 35.83 ± 2.59 | ^a 33.13 ± 0.75 | ^a 30.63 ± 1.02 | ^a 29.10 ± 2.06 |
| NTC | ^b 44.53 ± 1.30 | ^b 42.39 ± 0.52 | ^b 42.80 ± 0.61 | ^b 43.02 ± 0.98 |
| NTC + PHF (750 gm/ton) | ^b 42.74 ± 1.46 | ^b 43.09 ± 0.83 | ^b 41.63 ± 2.46 | ^b 42.26 ± 3.41 |
| NTC + PHF (1,000 gm/ton) | ^b 42.45 ± 0.84 | ^b 42.42 ± 1.00 | ^b 40.13 ± 0.95 | ^b 40.40 ± 1.78 |
| NTC + PHF (2,000 gm/ton) | ^b 44.72 ± 1.50 | ^b 42.01 ± 0.84 | ^b 39.71 ± 1.76 | ^b 39.81 ± 2.16 |

Values are expressed as mean ± SEM; *n* = 6; ^{a-b}Means bearing different superscripts were significantly different (*p* < 0.05) by One-way ANOVA followed by LSD using SPSS.

Table 7. Effect of PHF on cecal consistency score and cecal moisture content.

| Group | Cecal consistency score | Cecal moisture (gm/100 gm) |
|--------------------------|----------------------------|----------------------------|
| NC | ^a 1.00 ± 0.00 | ^a 73.70 ± 2.93 |
| NTC | ^b 4.00 ± 0.00 | ^b 87.16 ± 0.67 |
| NTC + PHF (750 gm/ton) | ^{b-c} 3.50 ± 0.22 | ^b 84.68 ± 2.60 |
| NTC + PHF (1,000 gm/ton) | ^c 3.00 ± 0.37 | ^b 83.26 ± 2.78 |
| NTC + PHF (2,000 gm/ton) | ^c 2.83 ± 0.48 | ^b 82.84 ± 2.19 |

Values are expressed as mean ± SEM; *n* = 6; ^{a-c}Means bearing different superscripts were significantly different (*p* < 0.05) by One-way ANOVA followed by LSD using SPSS.

Conclusion

In the present study, a wet litter broiler model was developed through the excessive feeding of MgCl₂ diet, which is characterized by the worsening of performance parameters and the emergence of problems associated with wet litter such as footpad lesion, high litter and cecal moisture, and poor cecal consistency. Supplementation with PHF ameliorated these problems and, therefore, it can be used as a natural alternative for the management of loose droppings and wet litter problems that arise during broiler production.

Acknowledgments

The authors expressed their deep gratitude to Dr. Muralidhar, Department of Animal Health Science for his technical assistance.

Conflict of interests

The authors declare no conflict of interest.

Authors' contribution

Saravanakumar Marimuthu performed the work, compiled, analyzed and interpreted the data, and drafted the manuscript, Brindhalakshmi Balasubramanian performed the work and reviewed the manuscript, Ramasamy Selvam planned the study, analyzed and interpreted the data, and proof read and reviewed the manuscript, and Prashanth D'Souza planned the study and reviewed the manuscript.

References

- [1] Stephenson AH, McCaskey TA, Ruffin BG. A survey of broiler litter composition and potential value as a nutrient resource. *Biol Waste* 1990; 34:1-9; [https://doi.org/10.1016/0269-7483\(90\)90139-J](https://doi.org/10.1016/0269-7483(90)90139-J)
- [2] Torok VA, Hughes RJ, Ophel-Keller K, Ali M, MacAlpine R. Influence of different litter materials on cecal microbiota colonization in broiler chickens. *Poult Sci* 2009; 88:2474-81; <https://doi.org/10.3382/ps.2008-00381>
- [3] Mayne RK, Else RW, Hocking PM. High litter moisture alone is sufficient to cause footpad dermatitis in growing turkeys. *Br Poult Sci* 2007; 48:538-45; <https://doi.org/10.1080/00071660701573045>
- [4] Abd El-Wahab A. Experimental studies on effects of diet composition (electrolyte contents), litter quality (type, moisture) and infection (coccidia) on the development and severity of foot pad dermatitis in young turkeys housed with or without floor heating. PhD Thesis, University of Veterinary Medicine, Hannover, Germany, 2011.
- [5] Weaver WD Jr, Meijerhof R. The effect of different levels of relative humidity and air movement on litter conditions, ammonia levels, growth, and carcass quality for broiler chickens. *Poult Sci* 1991; 70:746-55; <https://doi.org/10.3382/ps.0700746>
- [6] Mitran L, Harter-Dennis JM, Meisinger JJ. Determining the nitrogen budget and total ammoniacal nitrogen emissions from commercial broilers grown in environmental chambers. *J Appl Poult Res* 2008; 17:34-46; <https://doi.org/10.3382/japr.2006-00125>
- [7] Francesch M, Brufau J. Nutritional factors affecting excreta/litter moisture and quality. *World's Poult Sci J* 2004; 60:64-75; <https://doi.org/10.1079/WPS20035>
- [8] Montagne L, Pluske JR, Hampson DJ. A review of interactions between dietary fibre and the intestinal mucosa, and their consequences on digestive health in young non-ruminant animals. *Anim Feed Sci Technol* 2003; 108:95-117; [https://doi.org/10.1016/S0377-8401\(03\)00163-9](https://doi.org/10.1016/S0377-8401(03)00163-9)
- [9] Wu K, Hocking PM. Turkeys are equally susceptible to foot pad dermatitis from 1 to 10 weeks of age and foot pad scores were minimized when litter moisture was less than 30%. *Poult Sci* 2011; 90:1170-8; <https://doi.org/10.3382/ps.2010-01202>
- [10] Youssef IMI, Beineke A, Rohn K, Kamphues J. Effects of litter quality (moisture, ammonia, uric acid) on development and severity of foot pad dermatitis in growing turkeys. *Avian Dis* 2011; 55:51-8; <https://doi.org/10.1637/9495-081010-Reg.1>
- [11] Meluzzi A, Fabbri C, Folegatti E, Sirri F. Survey of chicken rearing conditions in Italy: Effects of litter quality and stocking density on productivity, foot dermatitis and carcass injuries. *Br Poult Sci* 2008; 49:257-64; <https://doi.org/10.1080/00071660802094156>
- [12] De Jong IC, Gunnink H, van Harn J. Wet litter not only induces footpad dermatitis but also reduces overall welfare, technical performance, and carcass yield in broiler chickens. *J Appl Poult Res* 2014; 23:51-8; <https://doi.org/10.3382/japr.2013-00803>
- [13] Ravindran V, Morel PC, Partridge GG, Hruba M, Sands JS. Influence of an *E. coli*-derived phytase on nutrient utilization in broiler starters fed diets containing varying concentrations of phytic acid. *Poult Sci* 2006; 85:82-9; <https://doi.org/10.1093/ps/85.1.82>

- [14] van der Hoeven-Hangoor E, Rademaker CJ, Paton ND, Verstegen MWA, Hendriks WH. Evaluation of free water and water activity measurements as functional alternatives to total moisture content in broiler excreta and litter samples. *Poult Sci* 2014; 93(7):1782–92; <https://doi.org/10.3382/ps.2013-03776>
- [15] Fairbrother JM, Nadeau E, Gyles CL. *Escherichia coli* in postweaning diarrhea in pigs: an update on bacterial types, pathogenesis, and prevention strategies. *Anim Health Res Rev* 2005; 6(1):17–39; <https://doi.org/10.1079/AHR2005105>
- [16] Casewell M, Friis C, Marco E, McMullin P, Phillips I. The European ban on growth-promoting antibiotics and emerging consequences for human and animal health. *J Antimicrob Chemother* 2003; 52:159–61; <https://doi.org/10.1093/jac/dkg313>
- [17] Hofacre CL, Beacorn T, Collett S, Mathis G. Using competitive exclusion, mannan-oligosaccharide and other intestinal products to control necrotic enteritis. *J Appl Poult Res* 2003; 12:60–4; <https://doi.org/10.1093/japr/12.1.60>
- [18] Etheridge RD, Seerley RW, Huber TL. The effect of diet on fecal moisture, osmolarity of fecal extracts, products of bacterial fermentation and loss of minerals in feces of weaned pigs. *J Anim Sci* 1984; 58:1403–11; <https://doi.org/10.2527/jas1984.5861403x>
- [19] Lee S, Britton WM. Magnesium toxicity—effect on phosphorus utilization by broiler chicks. *Poult Sci* 1980; 59:1989–94; <https://doi.org/10.3382/ps.0591989>
- [20] Vu MK, Nouwens MAG, Biemond I, Lamers C, Masclee AAM. The osmotic laxative magnesium sulphate activates the ileal brake. *Aliment Pharmacol Ther* 2000; 14:587–95; <https://doi.org/10.1046/j.1365-2036.2000.00746.x>
- [21] Schiller LR. Review article: the therapy of constipation. *Aliment Pharmacol Ther* 2001; 15:749–63; <https://doi.org/10.1046/j.1365-2036.2001.00982.x>
- [22] Galvez J, Crespo ME, Zarzuelo A, Dewitte P, Spiessens C. Pharmacological activity of a procyanidin isolated from *Sclerocarya birrea* bark—antidiarrheal activity and effects on isolated guinea-pig ileum. *Phytother Res* 1993; 7:25–8; <https://doi.org/10.1002/ptr.2650070108>
- [23] Uddin SJ, Shilpi JA, Alam SMS, Alamgir M, Rahman MT, Sarker SD. Antidiarrhoeal activity of the methanol extract of the barks of *Xylocarpus moluccensis* in castor oil- and magnesium sulphate-induced diarrhoea models in mice. *J Ethnopharmacol* 2005; 101:139–43; <https://doi.org/10.1016/j.jep.2005.04.006>
- [24] Antonisamy P, Kannan P, Ignacimuthu S. Anti-diarrhoeal and ulcer-protective effects of violacein isolated from *Chromobacterium violaceum* in Wistar rats. *Fundam Clin Pharmacol* 2009; 23:483–90; <https://doi.org/10.1111/j.1472-8206.2009.00701.x>
- [25] Ikarashi N, Ushiki T, Mochizuki T, Toda T, Kudo T, Baba K, et al. Effects of magnesium sulphate administration on aquaporin 3 in rat gastrointestinal tract. *Biol Pharm Bull* 2011; 34:238–42; <https://doi.org/10.1248/bpb.34.238>
- [26] Lee SR, Britton WM. Magnesium induced catharsis in broiler males proceeds via neural mechanisms. *Poult Sci* 1983; 62:1456.
- [27] Durlach J, Guiet-Bara A, Pages N, Bac P, Bara M. Magnesium chloride or magnesium sulfate: a genuine question. *Magnes Res* 2005; 18:187–92.
- [28] van der Hoeven-Hangoor E, Paton ND, van de Linde IB, Verstegen MWA, Hendriks WH. Moisture content in broiler excreta is influenced by excreta nutrient contents. *J Anim Sci* 2013; 91:5705–13; <https://doi.org/10.2527/jas.2013-6573>
- [29] Selvam R, Saravanakumar M, Suresh S, Chandrasekaran CV, D'Souza P. Evaluation of polyherbal formulation and synthetic choline chloride on choline deficiency model in broilers: implications on zootechnical parameters, serum biochemistry and liver histopathology. *Asian-Australas J Anim Sci* 2018; 31(11):1795–806; <https://doi.org/10.5713/ajas.18.0018>
- [30] Hashimoto S, Yamazaki K, Obi T, Takase K. Footpad dermatitis in broiler chickens in Japan. *J Vet Med Sci* 2011; 73:293–7; <https://doi.org/10.1292/jvms.10-0329>
- [31] Liu YX, Guo YM, Wang Z, Nie W. Effects of source and level of magnesium on catalase activity and its gene expression in livers of broiler chickens. *Arch Anim Nutr* 2007; 61:292–300; <https://doi.org/10.1080/17450390701432019>
- [32] Guo YM, Zhang GM, Yuan HM, Nie W. Effects of source and level of magnesium and vitamin E on prevention of hepatic peroxidation and oxidative deterioration of broiler meat. *Anim Feed Sci Technol* 2003; 107:143–50; [https://doi.org/10.1016/S0377-8401\(03\)00116-0](https://doi.org/10.1016/S0377-8401(03)00116-0)
- [33] Xing JH, Soffer EE. Adverse effects of laxatives. *Dis Colon Rectum* 2001; 44:1201–09; <https://doi.org/10.1007/BF02234645>
- [34] Saris NEL, Mervaala E, Karppanen H, Khawaja JA, Lewenstam A. Magnesium—An update on physiological, clinical and analytical aspects. *Clin Chim Acta* 2000; 294:1–26; [https://doi.org/10.1016/S0009-8981\(99\)00258-2](https://doi.org/10.1016/S0009-8981(99)00258-2)
- [35] McKeegan D. Foot pad dermatitis and hock burn in broilers: risk factors, aetiology and welfare consequences, 2010. <http://randd.defra.gov.uk/Default.aspx?Menu=Menu&Module=More&Location=None&Completed=0&ProjectID=14546> (Accessed April 2013)
- [36] Cengiz O, Hess JB, Bilgili SF. Effect of bedding type and transient wetness on footpad dermatitis in broiler chickens. *J Appl Poult Res* 2011; 20:554–60; <https://doi.org/10.3382/japr.2011-00368>
- [37] Masnindah M, Abdul RA, Awis QS, Sharida F. Effects of *Andrographis paniculata* and *Orthosiphon stamineus* supplementation in diets on growth performance and carcass characteristics of broiler chickens. *Int J Agric Biol* 2013; 15:897–902.
- [38] Lansky E, Shubert S, Neeman I. Pharmacological and therapeutic properties of pomegranate. In: Melgarejo-Moreno P, Martinez-Nicolas JJ, Martinez-Tome J (eds.). Production, processing and marketing of pomegranate in the Mediterranean region: advances in research and technology. CIHEAM-IAMZ, Zaragoza, Spain, pp 231–5, 2000.
- [39] Qnais EY, Elokda AS, Abu Ghalyun YY, Abdulla FA. Antidiarrheal activity of the aqueous extract of *Punica granatum* (Pomegranate) peels. *Pharm Biol* 2007; 45(9):715–20; <https://doi.org/10.1080/13880200701575304>
- [40] Gupta S, Yadava JNS, Tandon JS. Antisecretory (antidiarrhoeal) activity of Indian medicinal plants against *Escherichia coli* enterotoxin-induced secretion in rabbit and guinea pig ileal loop models. *Int J Pharmacogn* 1993; 31(3):198–204; <https://doi.org/10.3109/13880209309082942>
- [41] Gupta S, Choudhry MA, Yadava JNS, Srivastava V, Tandon JS. Antidiarrhoeal activity of diterpenes of *Andrographis paniculata* (Kal-Megh) against *Escherichia coli* enterotoxin in *in vivo* models. *Int J Crude Drug Res* 1990; 28(4):273–83; <https://doi.org/10.3109/13880209009082833>
- [42] Misar A, Bhagat R, Mujumdar AM. Antidiarrhoeal activity of *Acacia nilotica* Willd. bark methanol extract. *Hindustan Antibiot Bull* 2007; 49–50(1–4):14–20.
- [43] Bimlesh K, Kalyani D, Prashant T, Manoj S, Diwakar G. Evaluation of anti-diarrhoeal effect of aqueous and ethanolic extracts of fruit pulp of *Terminalia bellerica* in rats. *Int Drug Dev Res* 2010; 2(4):769–79.
- [44] Basu NK, Jayaswal SB. Amoebicidal activity of alkaloids *in vitro*. *Indian J Pharm* 1968; 30:289.
- [45] Mishra US, Mishra A, Kumari R, Murthy PN, Naik BS. Antibacterial activity of ethanol extract of *Andrographis paniculate*. *Indian J Pharm Sci* 2009; 71(4):436–8; <https://doi.org/10.4103/0250-474X.57294>
- [46] Sotohy SA, Sayed AN, Ahmed MM. Effect of tannin-rich plant (*Acacia nilotica*) on some nutritional and bacteriological parameters in goats. *Dtsch Tierarztl Wochenschr* 1997; 104(10):432–5.

- [47] Anindita D, Barua S, Das B. Pharmacological activities of Baheda (*Terminalia bellerica*): a review. *J Pharmacogn Phytochem* 2016; 5(1):194–7.
- [48] Anup B, Jamarkattel N, Shrestha A, Lamsal NK, Shakya S, Rajbhandari S. Evaluation of antioxidative and antidiabetic activity of bark of *Holarrhena pubescens* wall. *J Clin Diagn Res* 2014; 8(9):5–8.
- [49] Esteban CJ, Durban RF, Lopez-Argueta AS, Lopez MJ. A comparative analysis of response to vs. ORS + gelatin tannate pediatric patients with acute diarrhea. *Rev Esp Enferm Dig* 2009; 101:41–8.
- [50] Loeb H, Vandenplas Y, Wursch P, Guesry P. Tannin-rich carob pod for the treatment of acute-onset diarrhea. *J Pediatr Gastroenterol Nutr* 1989; 8:480–5; <https://doi.org/10.1097/00005176-198905000-00010>
- [51] van Ampting MT, Schonewille AJ, Vink C, Brummer RJ, van der Meer R, Bovee-Oudenhoven IM. Damage to the intestinal epithelial barrier by antibiotic pretreatment of salmonella-infected rats is lessened by dietary calcium or tannic acid. *J Nutr* 2010; 140:2167–72; <https://doi.org/10.3945/jn.110.124453>
- [52] Chen CH, Liu TZ, Chen CH, Wong CH, Chen CH, Lu FJ, et al. The efficacy of protective effects of tannic acid, gallic acid, ellagic acid, and propyl gallate against hydrogen peroxide-induced oxidative stress and DNA damages in IMR-90 cells. *Mol Nutr Food Res* 2007; 518:962–8; <https://doi.org/10.1002/mnfr.200600230>
- [53] Tikoo K, Tamta A, Ali IY, Gupta J, Gaikwad AB. Tannic acid prevents azidothymidine (AZT) induced hepatotoxicity and genotoxicity along with change in expression of PARG and histone H3 acetylation. *Toxicol Lett* 2008; 177:90–6; <https://doi.org/10.1016/j.toxlet.2007.12.012>
- [54] Darji VC, Deshpande SS, Bariya AH. Effects of methanolic extract of *Holarrhena antidysenterica* bark against experimentally induced inflammatory bowel disease in rats. *Int Res J Pharm* 2012; 3(9):152–4.
- [55] Solanki R, Madat D, Chauhan K, Adeshara SP. Analgesic activity of *Holarrhena antidysenterica* (Apocynaceae) bark. *Int J Pharma Phytochem Res* 2012; 2(4):5–7.
- [56] Mira AD, Ratnawatia J, Purwasih RW. Determination of total tannin of white and red rind pomegranate (*Punica granatum* L.) by colorimetry method using reagent 1, 10 Phenantroline. *Procedia Chem* 2014; 13:214–17; <https://doi.org/10.1016/j.proche.2014.12.030>